Impact of Climate Change on Plant Diseases

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Abstract

Climate change is profoundly altering the dynamics of plant diseases, with significant implications for global agriculture and ecosystem stability. This review explores how rising temperatures, shifts in precipitation patterns, and elevated atmospheric CO_2 levels influence the incidence, distribution, and severity of plant diseases based on recent studies. Global temperature rise expands the geographic range of pathogens, including fungi, bacteria, viruses and oomycetes, while changes in moisture availability creates favorable conditions for both drought tolerant and water-dependent pathogens. Elevated CO_2 may further affect plant-pathogen interactions by altering plant growth and resistance mechanisms. Additionally, climate-induced phenological shifts are leading to changes in the timing and frequency of disease outbreaks. Therefore, understanding these complex plant-pathogen interactions are essential for developing adaptive management strategies, importantly, breeding of climate-resilient crops, disease forecasting models, and integrated pest management practices. Proactive efforts are critical to mitigate the increasing risks of plant diseases with the rapidly changing climate, globally.

Keywords: Agriculture, Climate change, Disease epidemiology, Mitigation strategies, Plant diseases

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Introduction

Climate change refers to long-term alterations in temperature, precipitation, wind patterns, and other elements of Earth's climate system (IPCC, 2021). It is driven mainly by the emission of greenhouse gases from human activities such as burning fossil fuels and gases deforestation. These trap atmospheric heat, leading to rising temperatures global and shifting weather patterns. The consequences of human activities on world climate have reached an alarming state, posing critical threats to physical and socioeconomic structures. The globallyaveraged surface concentration of CO₂ reached 420.0 parts per million (ppm), methane 1,934 parts per billion (ppb) and nitrous oxide 336.9 ppb in 2023. These values are 151%, 265% and 125% of pre-industrial (before 1750)

levels, respectively (WMO, 2024). Carbon dioxide emissions, which account for the maximum proportion of greenhouse gases (Sathaye *et al.*, 2006), rose to 37.4 billion tons (Gt) in 2023 (IEA, 2024). The Intergovernmental Panel on Climate Change has projected that global temperatures will have a further increase of 1.5 °C between 2030 and 2052 (IPCC, 2014).

As mentioned by Mendelsohn (2009), agriculture is the most vulnerable sector to climate change due to its large scale and sensitivity to weather parameters, thereby causing huge economic impacts. While, pathogens and pests are major causes of crop losses and are responsible for estimated average global losses of 17.2 % to 30.0 % in staple crops (Savary *et al.*, 2019). The interaction between pathogen, host, and

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conditions. environmental the components of the disease-triangle, is the key in determining the disease incidence. Climate change has a great effect on all these three factors (Zayan, 2019) by intensifying and broadening the impact of plant diseases through alterations in environmental conditions that favour the plant-pathogen interactions. Outbreaks of plant diseases affect both environmental sustainability and global food security by reducing primary production and biodiversity worldwide (Fones et al., 2020). Climate change may facilitate plant infection in multiple ways, including altering pathogen evolution, changing hostpathogen interactions and facilitating the emergence of new strains of pathogens, which in turn can break down host-plant resistance (Velásquez et al., 2018). Burdon and Zhan (2020) have also indicated that the plant diseases might proliferate into new regions as a result of pathogens shift their geographic ranges due to climate change.

Given the complexity and variability of plant-pathogen interactions. understanding the mechanisms by which climate change influences plant diseases are critical for developing adaptive management strategies. Thus, this review focused on exploring the current understanding of how climate change affects plant disease dynamics, shifts focusing on in pathogen distribution, changes in disease severity, interactions between and the environmental disease stress and susceptibility. Additionally, we will address the growing threat of plant diseases under changing climatic conditions.

Effect of climate change on the development of plant diseases

Elevated temperature

The temperature elevation is a crucial factor in disease development and severity as proven from various studies in the past. As indicated by Kweku et al. (2019), in a disease development the critical aspects of the pathogen infection process, such as latency period, sporulation, inoculum dissemination. and host resistance dynamics, are controlled by temperature. While, the reproduction and survival of pathogens significantly influenced are bv temperature, which further influences the disease severity (Campbell and Madden, 1990). A recent study has shown that high temperatures encourage the growth and spread of plant diseases, particularly bacteria and fungi, while also retards host defensive systems (Devi et al., 2022). This impact of the temperature elevated in disease development and disease severity could be further analyzed with some examples from the past research works; wheat vellow rust started to appear in late December due to increased et al.. temperatures (Jindal 2012). Regarding the interaction between the rust fungus Puccinia striiformis and wheat, new pathogen races that are more aggressive in causing disease at higher temperatures have appeared since the year 2000 and have become more prevalent worldwide in only a few years (Hovmøller et al., 2008; Milus et al., 2009). Stefansson et al. (2013) found that the local adaptation to stressful, high temperature as well as mean annual temperature variation in addition to a high adaptive potential for growth rate at all experimental temperatures in most populations of Rhynchosporium commune. When rice plants are preexposed to heat stress, their resistance to the fungal pathogen Magnaporthe orvzae is weakened, leading to faster tissue necrosis, increased pathogen proliferation, and more severe disease symptoms (Onaga et al., 2016). In southern Germany, a northward shift of Cercospora beticola, leaf spot of sugar beet was due to increase in the annual mean temperature by 0.8-1 °C (Saxony et al., 2011). On the other hand, the temperature rises in the environment also alter the host resistance factor. As an example, in Arabidopsis thaliana, high temperature reduces expression of the immune receptor FLS2, which suppresses immunity to the pathogen Pseudomonas syringae pv. tomato (Janda et al., 2019). Thus, the ability of pathogens to adapt to higher temperatures over time, along with reduced immunity in plants under heat stress, has led to an increase in disease incidence at elevated temperatures.

It is important to remember that there are differences in the ideal temperature range for pathogen-host systems, as certain pathogens can be sensitive to high or low temperatures (Porras et al., 2023). For example, temperature above 30 °C reduces the ability of spores of the pathogen Microbotryum fungal lychnidis-dioicae, which causes а sterilizing anther-smut disease on the herbaceous plant Silene latifolia, to germinate, grow, and conjugate in vitro, whereas temperature between 26 °C and 31 °C are ideal for papaya ring spot virus (PRSV) to infect papaya (Mangrauthia al., 2009). et Xanthomonas oryzae pv. oryzae (Xoo), that fails to efficiently colonize rice xylem when the daytime temperature exceeds 35 °C (Horino et al., 1982). These evidences showed that each pathogen has its preferred temperature for host-pathogen interactions to cause disease.

A more virulent and better-adapted strain of a pathogen may emerge due to rising temperature (Cohen and Leach, 2020). For example, fusarium head blight of wheat has re-emerged in the USA, favored by warm weather at anthesis (Chakraborty and Newton, 2011). Previous studies showed that increased temperatures can worsen the intensity of various crop diseases, wheat rust (Puccinia including triticina), rice blast (Magnaporthe grisea), potato late blight infestans), and citrus (*Phytophthora* canker (Xanthomonas spp.) (Ahmed et al., 2024; Singh et al., 2023). Tomato plants are more susceptible to tomato vellow leaf curl virus during heat stress (Anfoka et al., 2016). In Central America, warmer temperatures contributed to severe epidemics of coffee rust in 2012–2013, by increasing infection rates. In East Africa, rising temperatures have allowed rust to spread to higher altitudes, while in Southeast Asia, increased temperature have worsened coffee rust outbreak (Koutouleas, 2023; Toniutti et al., 2017).

Elevated carbon dioxide level

Elevated CO₂ levels cause physiological changes in plant morphology such as increases in leaf size, leaf thickness, and number of leaves (Pritchard et al., 1999). A thick canopy leads to increased duration of leaf surface moisture, presenting a highly favorable environment for spore germination and penetration in foliar pathogens (Garrett et al., 2006). However, enhanced plant growth may also create denser canopies and microclimates that favor pathogen proliferation, particularly for foliar pathogens that thrive in humid environments (Melloy et al., 2010). For example, enhanced canopy density increases leaf surface moisture retention and creates a favorable environment for pathogens like Puccinia striiformis, the causative agent of stripe rust. This pathogen thrives in high humidity and cool temperatures, leading to severe outbreaks under such conditions (Chen, 2005). In potato fields, dense canopies can trap humidity, creating a moist microclimate favorable for *Phytophthora infestans*, the late blight pathogen, relies on wet foliage to germinate and infect host tissues, leading to significant yield losses during humid seasons (Fry, 2008).

Plant immune responses and hormone levels are influenced by atmospheric CO₂ level, which can affect the plantpathogen interaction. For example, increased basal expression of jasmonic acid-responsive genes under elevated CO₂ enhanced resistance to the necrotrophic leaf pathogen Botrytis cinerea, but reduced resistance to the hemi-biotrophic leaf pathogen Pseudomonas syringae pv. Tomato (Zhou et al., 2019). Reduction in the effectiveness of plant defense pathways under elevated CO₂ increased the susceptibility of wheat against the two major pathogens Zymoseptoria tritici and Fusarium graminearum that cause Septoria tritici blotch and Fusarium head blight, respectively (Váry et al., 2015). Elevated CO₂ levels increased the severity of powdery mildew on cucurbits caused by Sphaerotheca fuliginea (Khan and Rizvi, 2020).

Change in precipitation patterns

The change in precipitation patterns has diverse effects on the development of plant diseases. For instance, increased rainfall can increase the frequency and intensity of soil-borne and foliar diseases, such as root rot, damping-off, leaf spot, and blight (Lamichhane et al., 2023). Extended periods of leaf wetness due to increased rainfall increase the severity and rate of spread of soybean rust caused by Phakopsora pachyrhiziin (Narváez et al., 2010). In alfalfa, Phoma medicaginis is responsible for leaf spot and black stem diseases, which cause substantial vield reductions. The pathogen spreads to healthy plants rain-splashed through spores originating from infected plant debris (Lan et al., 2014). In groundnuts, wet weather promotes the spread of **Phaeoisariopsis** personata, the pathogen behind late leaf spot. This disease causes premature defoliation and leads to significant yield losses, particularly in areas with intense seasonal rainfall. Penet et al. (2014) found that direct rain splash dispersal being the dominant dispersal mechanism Colletotrichum of gloeosporioides conidia while re-splash from contaminated soil contributed less to dispersal.

Increased rainfall may also increase the risk of flooding and waterlogging, which can create anaerobic conditions, that favors the development of soilborne pathogens, such as *Pythium* spp. and *Phytophthora* spp. (Martínez-Arias et al., 2022). Decreased rainfall may increase the susceptibility of host plants to drought stress and attack of pathogens, including Fusarium spp. and Verticillium spp. (Maurya et al., 2022). Drier conditions may also reduce the effectiveness of certain biological control agents such as *Trichoderma* spp. and *Pseudomonas fluorescens*, that rely on moist conditions for their survival, leading to increased disease prevalence (Harvell et al., 2002). Pseudomonas fluorescens is a well-known biocontrol agent against foliar and root diseases. It suppresses pathogens by producing antibiotics like 2. 4diacetylphloroglucinol (DAPG) and siderophores that chelate iron. However, drier conditions limit the bacterium's ability to colonize the rhizosphere or leaf surface effectively, diminishing its ability to suppress such *Phytophthora* pathogens as al.. infestans (Landa et 2002). Precipitation plays a major role in the dispersal mechanisms and spore

germination of pathogens, as well as influencing plant disease management.

Change in humidity levels

The change in humidity levels is expected to have a significant impact on the development of plant diseases. Humidity significantly influences the formation and deposition of dew, providing a vital source of free water that facilitates the survival and infection processes of many plant pathogens (Nath, 2021). For example, higher humidity levels can increase the frequency and severity of foliar diseases such as leaf spot and fruit diseases such as anthracnose (Colletotrichum spp.), scab (Streptomyces scabies), and gray mold (Botrytis cinerea) (Ji et al., 2021; Maurya et al., 2022). Obanor et al. (2008) found that increased leaf wetness duration led to a higher number of Spilocaea oleagina conidia germination on olive leaves at all temperatures tested (5–25 °C). Increased humidity may also increase the risk of post-harvest diseases such as soft rot and stem end rot, which can cause significant losses transportation in storage and (Moradinezhad and Ranjbar, 2023). Infection rates by Sclerotinia sclerotiorum in lettuce (Mamo et al., 2021) and the stem rot pathogen Phytophthora sojae are higher under increased humidity (Tada et al., 2021).

High humidity significantly inhibits the accumulation and signaling of salicylic acid (SA), a key defense hormone in Arabidopsis. NPR1, an SA receptor and co-activator of SA-responsive genes, is less ubiquitinated and has a reduced affinity for promoter binding under high humidity. which increase disease susceptibility in Arabidopsis. The down regulation of the cellular ubiquitination machinery, particularly the Cullin 3based E3 ubiquitin ligase, impairs NPR1 protein ubiquitination in these conditions (Yao et al., 2023). Humidity

significantly influences plant disease outbreaks, with high ambient humidity enhancing rice blast development caused by *Magnaporthe oryzae*. Study increased showed that humidity promotes conidial germination and appressorium formation, boosting the pathogen's virulence. High humidity suppresses ethylene biosynthesis and signaling, which are critical for basal resistance in rice. These findings highlight that humidity compromises rice resistance by reducing ethylene pathway activation (Qiu et al., 2022). Decreased humidity may reduce the incidence and spread of some diseases, such as rust and smut, but may also increase the susceptibility of host plants to water stress and other diseases such as wilt and canker (Jeger, 2022). A study by Milod et al. (2021) revealed that low relative humidity (20–40 %) and high temperatures (30 °C or above) disease inhibited progression of cucumber powdery mildew caused by cichoracearum. Humidity Ervsiphe level affects plant diseases by pathogen activity promoting and suppressing plant defenses at high levels, while low humidity inhibits some pathogens but increases stress susceptibility.

Extreme weather conditions

Severe weather conditions cause physical damage, physiological stress, and biochemical changes in host plants, and it is one of the main environmental factors influencing plant diseases (Singh et al., 2023). Extreme weather conditions increase the susceptibility of host plants to diseases. For instance, storms can cause physical wounds on host plants, serving as entry point for pathogens causing various diseases such crown gall, rust and Aphidas transmitted viral diseases (Bastas, 2022). Extreme weather events can help spread of pathogens to new locations, as for the case of soybean rust, which was

introduced from Brazil into the United States by a hurricane (Fones *et al.*, 2020). *Fusarium oxysporum* f. sp. *cubense* Tropical Race 4, a new race of the causal agent of Fusarium wilt of banana, may have been spread in Mozambique by a cyclone, increasing the risk of further spread to adjacent farmlands and neighboring countries in Africa (García-Bastidas *et al.*, 2014).

Flooding can also rapidly spread many pathogens among adjacent farms and regions. Pathogens with flagella or swimming stages, such as the oomycete pathogen Phytophthora, and Ralstonia solanacearum are able to disperse rapidly through flooded regions to new susceptible plants (Browne et al., 2021; Feng et al., 2017). Heat waves and cold snaps may induce heat and oxidative stress in plants, compromising their photosynthesis and respiration and rendering them more susceptible to diseases such as leaf spot, and black rot (Tanveer et al., 2023). In addition, hail damage to protective layers such as the epidermis and cuticle can make plants more vulnerable to diseases like bacterial spot and downy mildew (Khadiri et al., 2023). These studies showed that extreme weather events significantly influence plant diseases by causing physical damage, spreading pathogens, and increasing plant susceptibility through stress and weakened defenses. Events like storms, temperature floods. and extremes facilitate pathogen dispersal and infection, intensifying the risk of disease outbreaks from one geographic region to another.

Adaptation Strategies for Agriculture

Breeding for disease-resistant crops

A key adaptation strategy to combat plant diseases under climate change is the development of disease-resistant crop varieties. Recently, the CRISPR/Cas9 system has been utilized to enhance disease resistance among different crops such as rice, cacao, wheat, tomato, and grapes (Ahmad et al., 2020; Brooks et al., 2014). This system allows for precise genome editing of various organisms via RNAguided DNA endonuclease activity. In soybean, Nagy et al. (2021) utilized CRISPR/Cas9 to develop a novel chromosome rearrangement method. This approach engineered two NLR gene clusters, Rpp1-like, conferring resistance to Phakopsora pachyrhizi, and Rps1, associated with resistance to Phytophthora sojae. This technique demonstrated a promising strategy for disease enhancing resistance in soybean. In the cacao tree (Theobroma cacao), genome editing was conducted using Agrobacterium-mediated

transient transformation to introduce CRISPR/Cas9 components that targeted non-expressor pathogenesis-related 3 (TcNPR3), a suppressor of the defense response, into cacao leaves and cotyledon cells. The TcNPR3 deleted leaves exhibited up-regulated expression of defense genes, such as pathogenesis related (PR) genes, and enhanced resistance to the black pod disease caused by Phytophthora tropicalis (Fister et al., 2018).

Beyond genome editing in crops, editing the genomes of fungal and oomycete pathogens can also provide new strategies for plant disease management (Paul et al., 2021). The CRISPR/Cas9 method was used for directed mutagenesis targeting pyrimidine biosynthesis and tryptophan biosynthesis genes (pyr4 and TrpE) of biotrophic plant pathogen Claviceps purpurea. The TrpE mutants showed no infection to rye plants because of a reduction of the plant hormone auxin, normallv which is synthesized by C. purpurea in Trp-dependent and Trp-independent biosynthetic pathways, ultimately affecting the ability of the fungus to colonize rye plants (Králová et al., 2021). Homozygous PpalEPIC8 mutants of the papaya pathogen *Phytophthora* palmivora were created using the CRISPR/Cas9 system, and it was observed that these mutants showed decreased ability to cause disease in This papaya fruits. reduction in pathogenicity occurred because the deletion of the cysteine protease inhibitor *PpalEPC8* led to a diminished suppression of papain, a cysteine protease in papaya that plays a role in defending against plant pathogens (Gumtow et al., 2018).

These methods can create crops that are more resilient to the changing pathogen pressures brought about by climate Panwar et (2018)shifts. al. demonstrates that generating transgenic wheat plants expressing RNAi-inducing transgenes to silence essential genes in rust fungi (Puccinia triticina) can provide effective disease resistance, thus opening an alternative way for developing rust-resistant crops. Tiwari et al. (2017) enhanced the resistance of rice to the sheath blight disease by down-regulating pathogenicity genes of Rhizoctonia solani using host-delivered RNAi. Ding et al. (2021) found that the appressorium-related gene Sscnd1 is required for cell integrity and full virulence in Sclerotinia sclerotiorum and that Sclerotinia stem rot can be controlled by expressing the silencing constructs of Sscnd1 in host plants. These techniques pave the way for breeding disease-resistant crops in order to overcome challenges from climate change in agriculture.

Improved forecasting and monitoring systems

Predictive modeling and disease forecasting tools have become essential in anticipating the emergence of plant

climate diseases under varving conditions. Climate-based forecasting systems help in the early detection of disease outbreaks, enabling timely interventions (Elad and Pertot, 2014). Optical techniques, such as RGB multi- and hyperspectral imaging. sensors, thermography, or chlorophyll fluorescence. have proven their potential in automated, objective, and reproducible detection systems for the identification and quantification of plant diseases at early time points in (Mahlein, epidemics 2016). Chlorophyll Fluorescence Imaging (CFI) is used to detect disruptions in photosynthesis. Chlorophyll Fluorescence Imaging has proven effective for identifying diseases like Xanthomonas infections in Arabidopsis at early stages. This technique relies on changes in fluorescence emission caused by stress, enabling a detailed study of pathogen effects (Sapoukhina et al., 2022). Oerke et al. (2011) and Belin et *al.* (2013) analyzed apple trees suffering from apple scab. In that case, the detection of infection, as well as differences in the virulence of several Venturia inaequalis isolates infecting apple trees, were detected more accurately by thermography. The thermal response was presymptomatic and consisted of spots of decreased temperature due to the subcuticular growth of V. inaequalis.

There has been a substantial effort to develop smart agricultural tools to help address the uncertainty surrounding plant disease epidemics through computational decision support systems (DSSs) (Zhai et al., 2020). AgroClimate is one of the example of a DSS designed to tackle weather- and climate-based risks in agriculture (Fraisse et al., 2016). AgroClimate provides several tools to assist farmers with plant disease management. The strawberry (Pavan et al., 2011), blueberry (Gama et al.,

2021), and citrus advisory systems (Perondi *et al.*, 2020) couple current and forecast weather data with disease models to predict risks and recommend fungicide applications when environmental conditions are favorable for disease development.

Cultural practices and agronomic adjustments

Agronomic adjustments, such as crop rotation, intercropping, soil tillage, altered planting dates, and liming and irrigation are important adaptation strategies to prevent or reduce the risk of diseases (Fry, 2012). These practices reduce the environmental can conditions favorable for pathogen proliferation. Enhanced diversity in cropping systems may reduce the risks of diseases that otherwise, in would monoculture. become more severe due to climate change. For example, Hannukkala et al. (2007) concluded that increased and earlier occurrence of late blight (Phytophthora infestans) epidemics in potato were probably associated with both climate change and lack of rotation.

Mitigation Strategies

Reducing greenhouse gas emissions from agriculture

Mitigation strategies aim to address the root cause of climate change by reducing greenhouse gas (GHG) emissions from agricultural practices. Agricultural activities, such as the use of synthetic fertilizers and livestock production, significantly contribute to the global GHG emissions (Smith et al., 2014). By implementing sustainable agricultural practices, such as organic farming, conservation agriculture, and the use of bio-fertilizers, farmers can reduce emissions and minimize the long-term impacts of climate change on plant diseases (Lal, 2004).

Carbon sequestration through agroforestry

Agroforestry, which integrates trees into agricultural landscapes, provides a dual benefit in both climate mitigation management. and disease Trees sequester atmospheric carbon, reducing the overall greenhouse gas burden (Jose, 2009). At the same time, agroforestry systems promote biodiversity, which can suppress the spread of plant diseases by creating more complex ecosystems that limit pathogen proliferation et al., 2000). Vertical (Schroth stratification of soil carbon stock under agroforestry is one of the options for long-term carbon sequestration and mitigating GHG emissions (Mishra et al., 2024).

Integrated pest management

Integrated Pest Management (IPM) is a holistic approach that combines biological, cultural, and chemical tools to manage plant diseases in a sustainable manner. Integrated Pest Management reduces the reliance on chemical pesticides, which are known to contribute to greenhouse gas emissions and can exacerbate environmental issues (Lamichhane et al., 2016). By incorporating climate-resilient practices, IPM can minimize disease outbreaks while promoting environmental sustainability.

Biotechnological approaches for climate-smart agriculture

Biotechnology, including genetic engineering and the use of microbial inoculants as biocontrol agents, offers promising solutions for managing plant diseases climate under stress. Genetically engineered drought-tolerant maize varieties, such as MON 87460 (developed by Monsanto), utilize genes like cold shock protein B (cspB) from Bacillus subtilis. These maize varieties are widely adopted in sub-Saharan Africa to mitigate yield losses due to water scarcity (Nemali *et al.*, 2015).

Recent advances in plant-microbiome research have shown that manipulating rhizosphere microbiome the can enhance plant resistance to both biotic and abiotic stresses (Compant et al., 2019). Targeting Induced Local Lesions in Genomes (TILLING) is a reverse genetic technique based on chemical induced mutagenesis coupled with a sensitive DNA screening-technique (Slade and Knauf, 2005) which allows the discovery of rare mutations in populations. Barley mutants were generated by TILLING to study the nucleotide variations in the eral (enhanced response to ABA1) gene (Daszkowska-Golec et al., 2018), which is differently regulated under drought tolerance in several species including wheat and soybean (Manmathan et al., 2013; Ogata et al., 2017).

Extremophiles are the microorganisms thriving in extreme environments apparent with harsh conditions such as thermophilic (high-temperature), psychrophilic temperature), (low halophilic (hypersaline environment), acidophilic (low pH), alkaliphilic (high pH), piezophilic (high pressure/deep radiophilic (high radiation). sea). xerophilic (no water), anaerobes (absence of oxygen), and snottite (in caves), and where normal life does not exist (Passarini et al., 2022). This can be utilized to combat complications faced by the agriculture sector in response to climate change as these genes which are responsible for their superior character can be incorporated into plant genome in order to produce stress resistant crop varieties.

Genome selection is an innovative tool in plant breeding to speed up crop improvement and high-throughput phenotyping to identify climateresilient genotypes for future breeding (Tadesse *et al.*, 2019). Climate-smart potato clones have been developed which showed high resistance to drought and heat stresses without yield penalty. The conditions given to these clones were 2-3 °C increases in temperature and about 15-20% lower precipitation under the scenario of climate change (Parker *et al.*, 2019).

Challenges and future directions

Although various adaptation and mitigation strategies show promise, several challenges remain. Climate projections change are inherently uncertain, making it difficult to predict the exact nature and magnitude of future plant disease outbreaks. Moreover, the global disparity in technological and financial resources can hinder the implementation climate-resilient of agricultural practices, particularly in developing nations (Nelson et al., 2009). Collaborative efforts between governments, research institutions, and farmers will be necessary to develop context-specific solutions that address these disparities and enhance global food security. Further research is needed to understand the complex interactions between climate change, physiology, and pathogen plant behavior. Integrating knowledge from plant pathology, climate science, and agricultural economics will provide a more comprehensive framework for tackling plant disease epidemics in a changing climate.

Conclusion

The impact of climate change on plant diseases is a complex and multifaceted issue that poses significant risks to agriculture and ecosystems. Changes in temperature, precipitation, and atmospheric CO_2 levels influence the biology of both plants and pathogens, potentially increasing the prevalence and severity of plant diseases. To address these challenges, proactive measures that integrate climate change considerations into plant disease management practices are necessary. Continued research and collaboration are critical for developing resilient agricultural systems that can withstand the increasing pressures of climateinduced plant diseases.

References

Ahmad, S., Wei, X., Sheng, Z., Hu, P. and Tang, S. (2020). CRISPR/Cas9 for development of disease resistance in plants: recent progress, limitations and future prospects. *Briefings in Functional Genomics* 19(1):26-39.

Ahmed, N., Alam, M., Saeed, M., Ullah, H., Junaid, M., Kanwal, M. and Ahmed, S. (2024). Role of Plants in Managing Diseases. In *Ethnic Knowledge and Perspectives of Medicinal Plants* (pp. 579-604). Apple Academic Press.

Anfoka, G., Moshe, A., Fridman, L., Amrani, L., Rotem, O.R., Kolot, M., Zeidan, M., Czosnek, H. and Gorovits, R. (2016). Tomato yellow leaf curl virus infection mitigates the heat stress response of plants grown at high temperatures. *Scientific Reports* 6(1):19715.

Bastas, K.K. (2022). Impact of climate change on food security and plant disease. In *Microbial Biocontrol: Food Security and Post Harvest Management: Volume 2* (pp. 1-22). Cham: Springer International Publishing.

Belin, É., Rousseau, D., Boureau, T. and Caffier, V. (2013). Thermography versus chlorophyll fluorescence imaging for detection and quantification of apple scab. *Computers and electronics in agriculture* 90:159-163.

Brooks, C., Nekrasov, V., Lippman, Z.B. and Van Eck, J. (2014). Efficient gene editing in tomato in the first generation using the clustered regularly interspaced short palindromic repeats/CRISPR-associated9 system. *Plant physiology* 166(3):1292-1297.

Browne, G.T., Hasey, J.K., Ott, N.J., Forbes, H., Arnold, K. and Milliron, L. (2021). Flooding by California rivers results in walnut scion infections by species of *Phytophthora*. *Plant Health Progress* 22(3):368-373.

Burdon, J.J. and Zhan, J. (2020). Climate change and disease in plant communities. *PLoS biology* 18(11):e3000949.

Campbell, C.L. and Madden, L.V. (1990). *Introduction to plant disease epidemiology* (pp. xvii+-532).

Chakraborty, S. and Newton, A.C. (2011). Climate change, plant diseases and food security: an overview. *Plant pathology* 60(1):2-14.

Chen, X.M. (2005). Epidemiology and control of stripe rust [*Puccinia striiformis* f. sp. *tritici*] on wheat. *Canadian journal of plant pathology* 27(3):314-337.

Cohen, S.P. and Leach, J.E. (2020). High temperature-induced plant disease susceptibility: more than the sum of its parts. *Current opinion in plant biology* 56:235-241.

Compant, S., Samad, A., Faist, H. and Sessitsch, A. (2019). A review on the plant microbiome: ecology, functions, and emerging trends in microbial application. *Journal of advanced research* 19:29-37.

Daszkowska-Golec, A., Skubacz, A., Sitko, K., Słota, M., Kurowska, M. and Szarejko, I. (2018). Mutation in barley ERA1 (Enhanced Response to ABA1) gene confers better photosynthesis efficiency in response to drought as revealed by transcriptomic and physiological analysis. *Environmental* and *Experimental Botany* 148:12-26.

Devi, R., Kaur, T., Kour, D., Yadav, A., Yadav, A.N., Suman, A., Ahluwalia, A.S. and Saxena, A.K. (2022). Minerals solubilizing and mobilizing microbiomes: A sustainable approach for managing minerals' deficiency in agricultural soil. *Journal of Applied Microbiology* 133(3):1245-1272.

Ding, Y., Chen, Y., Yan, B., Liao, H., Dong, M., Meng, X., Wan, H. and Qian, W. (2021). Host-induced gene silencing of a multifunction gene Sscnd1 enhances plant resistance against *Sclerotinia sclerotiorum. Frontiers in microbiology* 12:693334.

Elad, Y. and Pertot, I. (2014). Climate change impacts on plant pathogens and plant diseases. *Journal of Crop Improvement* 28(1):99-139.

Feng, S., Shu, C., Wang, C., Jiang, S. and Zhou, E. (2017). Survival of *Rhizoctonia solani* AG-1 IA, the causal agent of rice sheath blight, under different environmental conditions. *Journal of Phytopathology* 165(1):44-52.

Fister, A.S., Landherr, L., Maximova, S.N. and Guiltinan, M.J. (2018). Transient expression of CRISPR/Cas9 machinery targeting TcNPR3 enhances defense response in Theobroma cacao. *Frontiers in plant science* 9:268.

Fones, H.N., Bebber, D.P., Chaloner, T.M., Kay, W.T., Steinberg, G. and Gurr, S.J. (2020). Threats to global food security from emerging fungal and oomycete crop pathogens. *Nature Food* 1(6):332-342.

Fraisse, C., Andreis, J.H., Borba, T., Cerbaro, V., Gelcer, E., Pavan, W., Pequeno, D., Perondi, D., Shen, X., Staub, C. and Uryasev, O. (2016). AgroClimate-Tools for managing climate risk in agriculture. *Agrometeoros* 24(1):121-129.

Fry, W. (2008). *Phytophthora infestans*: the plant (and R gene) destroyer. *Molecular plant pathology*, 9(3):385-402.

Fry, W.E. (2012). *Principles of plant disease management*. Academic Press.

Gama, A.B., Cordova, L.G., Rebello, C.S. and Peres, N.A. (2021). Validation of a decision support system for blueberry anthracnose and fungicide sensitivity of *Colletotrichum gloeosporioides* isolates. *Plant Disease* 105(6):1806-1813.

García-Bastidas, F., Ordóñez, N., Konkol, J., Al-Qasim, M., Naser, Z., Abdelwali, M., Salem, N., Waalwijk, C., Ploetz, R.C. and Kema, G.H.J. (2014). First report of *Fusarium oxysporum* f. sp. *cubense* tropical race 4 associated with Panama disease of banana outside Southeast Asia. *Plant Disease* 98(5):694-694.

Garrett, K.A., Dendy, S.P., Frank, E.E., Rouse, M.N. and Travers, S.E. (2006). Climate change effects on plant disease: genomes to ecosystems. *Annu. Rev. Phytopathol.* 44(1):489-509.

Gumtow, R., Wu, D., Uchida, J. and Tian, M. (2018). A *Phytophthora palmivora* extracellular cystatin-like protease inhibitor targets papain to contribute to virulence on papaya. *Molecular Plant-Microbe Interactions* 31(3):363-373.

Hannukkala, A.O., Kaukoranta, T., Lehtinen, A. and Rahkonen, A. (2007). Late-blight epidemics on potato in Finland, 1933–2002; increased and earlier occurrence of epidemics associated with climate change and lack of rotation. *Plant pathology* 56(1):167-176. Harvell, C.D., Mitchell, C.E., Ward, J.R., Altizer, S., Dobson, A.P., Ostfeld, R.S. and Samuel, M.D. (2002). Climate warming and disease risks for terrestrial and marine

biota. Science 296(5576):2158-2162.

Horino, O., Mew, T.W. and Yamada, T. (1982). The effect of temperature on the development of bacterial leaf blight on rice. *Japanese Journal of Phytopathology* 48(1):72-75.

Hovmøller, M.S., Yahyaoui, A.H., Milus, E.A. and Justesen, A.F. (2008). Rapid global spread of two aggressive strains of a wheat rust fungus. *Molecular ecology* 17(17):3818-3826.

Intergovernmental Panel on Climate Change. (2014). In: Field CB, Barros VR, Dokken DJ, Mach KJ, Mastrandrea MD, Bilir TE, Chatterjee M, Ebi KL, Estrada YO, Genova RC, Girma B, Kissel ES, Levy AN, MacCracken S. Mastrandrea PR. White LL (eds) Climate change 2014: impacts. adaptation, and vulnerability. Part A: global and sectoral aspects. I. Cambridge Press. University Cambridge/New York, p. 1132.

Intergovernmental Panel on Climate Change. (2021). *Climate change 2021: The physical science basis*. <u>https://www.ipcc.ch/report/ar6/wg1/.</u>

International Energy Agency. (2024). CO₂ Emissions in 2023, Available from <u>https://iea.blob.core.windows.net/assets</u> /33e2badc-b839-4c18-84ce-

f6387b3c008f/CO

<u>2Emissionsin2023.pdf</u> [18 November 2024].

Janda, M., Lamparová, L., Zubíková, A., Burketová, L., Martinec, J. and Krčková, Z. (2019). Temporary heat stress suppresses PAMP-triggered immunity and resistance to bacteria in Arabidopsis thaliana. Molecular plant pathology 20(7):1005-1012.

Jeger, M.J. (2022). The impact of climate change on disease in wild plant populations and communities. *Plant Pathology* 71(1):111-130.

Ji, T., Salotti, I., Dong, C., Li, M. and Rossi, V. (2021). Modeling the effects of the environment and the host plant on the ripe rot of grapes, caused by the *Colletotrichum* species. *Plants* 10(11):2288.

Jindal, M.M., Mohan, C. and Pannu, P.P.S. (2012). Status of stripe rust of wheat in Punjab during 2011–12 season. *Proceedings of brain storming session. Department of Plant Pathology PAU, Ludhiana* 56.

Jose, S. (2009). Agroforestry for ecosystem services and environmental benefits: an overview (pp. 1-10). Springer, Dordrecht.

Khadiri, M., Boubaker, H., Askarne, L., Ezrari, S., Radouane, N., Farhaoui, A., El Hamss, H., Tahiri, A., Barka, E.A. and Lahlali, R. (2023). *Bacillus cereus* B8W8 an effective bacterial antagonist against major postharvest fungal pathogens of fruit. *Postharvest Biology and Technology* 200:112315.

Khan, M.R. and Rizvi, T.F. (2020). Effect of elevated levels of CO_2 on powdery mildew development in five cucurbit species. *Scientific Reports* 10(1):4986.

Koutouleas, A. (2023). Coffee leaf rust: wreaking havoc in coffee production areas across the tropics. *Plant health cases* (2023):phcs20230005.

Králová, M., Bergougnoux, V. and Frébort, I. (2021). CRISPR/Cas9 genome editing in ergot fungus *Claviceps purpurea. Journal of Biotechnology* 325:341-354. Kweku, D.W., Bismark, O., Maxwell, A., Desmond, K.A., Danso, K.B., Oti-Mensah, E.A., Quachie, A.T. and Adormaa, B.B. (2018). Greenhouse effect: greenhouse gases and their impact on global warming. *Journal of Scientific research and reports* 17(6):1-9.

Lal, R. (2004). Soil carbon sequestration to mitigate climate change. *Geoderma* 123(1-2):1-22.

Lamichhane, J.R., Barbetti, M.J., Chilvers, M.I., Pandey, A.K. and Steinberg, C. (2023). Exploiting root exudates to manage soil-borne disease complexes in a changing climate. *Trends in Microbiology* 32(1):27-37.

Lamichhane, J.R., Dachbrodt-Saaydeh, S., Kudsk, P. and Messéan, A. (2016). Toward a reduced reliance on conventional pesticides in European agriculture. *Plant Disease* 100(1):10-24.

Lan, Y., Zhou, W., Duan, T., Li, Y., Matthew, C. and Nan, Z. (2024). Alfalfa Spring Black Stem and Leaf Spot Disease Caused by *Phoma medicaginis*: Epidemic Occurrence and Impacts. *Microorganisms* 12(7):1279.

Landa. B.B.. Mavrodi. 0.V.. McSpadden Raaijmakers, J.M., Gardener, B.B., Thomashow, L.S. and Weller, D.M. (2002). Differential ability genotypes 2. of of 4diacetylphloroglucinol-producing Pseudomonas fluorescens strains to colonize the roots of pea plants. Applied Environmental and Microbiology 68(7):3226-3237.

Mahlein, A.K. (2016). Plant disease detection by imaging sensors–parallels and specific demands for precision agriculture and plant phenotyping. *Plant disease* 100(2):241-251.

Mamo, B.E., Eriksen, R.L., Adhikari, N.D., Hayes, R.J., Mou, B. and Simko, I. (2021). Epidemiological characterization of lettuce drop (*Sclerotinia* spp.) and biophysical features of the host identify soft stem as a susceptibility factor. *PhytoFrontiers*TM 1(3):182-204.

Mangrauthia, S.K., Singh Shakya, V.P., Jain, R.K. and Praveen, S. (2009). Ambient temperature perception in papaya for papaya ringspot virus interaction. *Virus genes* 38:429-434.

Manmathan, H., Shaner, D., Snelling, J., Tisserat, N. and Lapitan, N. (2013). Virus-induced gene silencing of *Arabidopsis thaliana* gene homologues in wheat identifies genes conferring improved drought tolerance. *Journal of experimental botany* 64(5):1381-1392.

Martínez-Arias, C., Witzell, J., Solla, A., Martin, J.A. and Rodríguez-Calcerrada, J. (2022). Beneficial and pathogenic plant-microbe interactions during flooding stress. *Plant, Cell & Environment* 45(10):2875-2897.

Maurya, M.K., Yadav, V.K., Singh, S.P., Jatoth, R., Singh, H.K. and Singh, D. (2022). Impact of Climate Change on Diseases of Crops and Their Management—A Review. *Journal of Agricultural Science and Technology B* 12(1):1-15.

Melloy, P., Hollaway, G., Luck, J.O., Norton, R.O.B., Aitken, E. and Chakraborty, S. (2010). Production and fitness Fusarium of pseudograminearum inoculum at elevated carbon dioxide in FACE. Global Change Biology 16(12):3363-3373.

Mendelsohn, R. (2009). The impact of climate change on agriculture in developing countries. *Journal of natural resources policy research* 1(1):5-19.

Milod, N., Saad, G. and Khalifa, H.A. (2021). Effect of temperature and relative humidity on conidial germination of the causal agent of cucumber powdery mildew. *Journal of International Medical Research and Health Sciences* 1:15-25.

Milus, E.A., Kristensen, K. and Hovmøller, M.S. (2009). Evidence for increased aggressiveness in a recent widespread strain of *Puccinia striiformis* f. sp. tritici causing stripe rust of wheat. *Phytopathology* 99(1):89-94.

Mishra, A.K., Das, R., Nath, D., Kumari, V., Mishra, S., Biswal, B. and Tyagi, R. (2024). Evidence-Based Agroforestry Systems as Climate-Resilient Farming Practices for Improving Livelihood and Sustainability in India. In Sustainable Management and Conservation of Environmental Resources in India (pp. 235-270). Apple Academic Press.

Moradinezhad, F. and Ranjbar, A. (2023). Advances in postharvest diseases management of fruits and vegetables: A review. *Horticulturae* 9(10):1099.

Nagy, E.D., Stevens, J.L., Yu, N., Hubmeier, C.S., LaFaver, N., Gillespie, M., Gardunia, B., Cheng, Q., Johnson, S., Vaughn, A.L. and Vega-Sanchez, M.E. (2021). Novel disease resistance gene paralogs created by CRISPR/Cas9 in soy. *Plant Cell Reports* 40:1047-1058.

Narváez, D.F., Jurick, W.M., Marois, J.J. and Wright, D.L. (2010). Effects of surface wetness periods on development of soybean rust under field conditions. *Plant disease* 94(2):258-264.

Nath, S. (2021). Dew as Source of Emerging Contaminants in Agricultural System. *Sustainable* Agriculture *Reviews 50: Emerging Contaminants in Agriculture*, pp.61-78.

Nelson, G.C., Rosegrant, M.W., Koo, J., Robertson, R., Sulser, T., Zhu, T., Ringler, C., Msangi, S., Palazzo, A., Batka. M. and Magalhaes, M. (2009). Climate change: Impact on agriculture and costs of adaptation (Vol. International 21). Food Policy Research Institute.

Nemali, K.S., Bonin, C., Dohleman, F.G., Stephens, M., Reeves, W.R., Nelson, D.E., Castiglioni, P., Whitsel, J.E., Sammons, B., Silady, R.A. and Anstrom, D. (2015). Physiological responses related to increased grain yield under drought in the first biotechnology-derived drought-tolerant maize. *Plant, Cell & Environment* 38(9):1866-1880.

Obanor, F.O., Walter, M., Jones, E.E. and Jaspers, M.V. (2008). Effect of temperature, relative humidity, leaf wetness and leaf age on *Spilocaea oleagina* conidium germination on olive leaves. *European Journal of Plant Pathology* 120:211-222.

Oerke, E.C., Fröhling, P. and Steiner, U. (2011). Thermographic assessment of scab disease on apple leaves. *Precision agriculture* 12:699-715.

Ogata, T., Nagatoshi, Y., Yamagishi, N., Yoshikawa, N. and Fujita, Y. (2017). Virus-induced down-regulation of GmERA1A and GmERA1B genes enhances the stomatal response to abscisic acid and drought resistance in soybean. *PLoS One* 12(4):e0175650.

Onaga, G., Wydra, K.D., Koopmann, B., Séré, Y. and von Tiedemann, A. (2016). Elevated temperature increases in planta expression levels of virulence related genes in *Magnaporthe oryzae* and compromises resistance in *Oryza sativa* cv. Nipponbare. *Functional Plant Biology* 44(3):358-371. Panwar, V., Jordan, M., McCallum, B. and Bakkeren, G. (2018). Host-induced silencing of essential genes in *Puccinia triticina* through transgenic expression of RNA i sequences reduces severity of leaf rust infection in wheat. *Plant biotechnology journal* 16(5):1013-1023.

Parker, M.L., Low, J.W., Andrade, M., Schulte-Geldermann, E. and Andrade-Piedra, J. (2019). Climate change and seed systems of roots, tubers and bananas: the cases of potato in Kenya and Sweetpotato in Mozambique. *The climate-smart agriculture papers: Investigating the Business of a productive, resilient and Low emission future* 99-111.

Passarini, M.R.Z., Duarte, A.W.F., Rosa, L.H., de Oliveira, V.M. and Ottoni, J.R. (2022). Extremofuels: Production of biofuels by extremophile microbes as an alternative to avoid climate change effects. In *Microbiome under Changing Climate* (pp. 237-256). Woodhead Publishing.

Paul, N.C., Park, S.W., Liu, H., Choi, S., Ma, J., MacCready, J.S., Chilvers, M.I. and Sang, H. (2021). Plant and fungal genome editing to enhance plant disease resistance using the CRISPR/Cas9 system. *Frontiers* in *Plant Science* 12:700925.

Pavan, W., Fraisse, C.W. and Peres, N.A. (2011). Development of a webbased disease forecasting system for strawberries. *Computers* and *electronics in agriculture* 75(1):169-175.

Penet, L., Guyader, S., Pétro, D., Salles, M. and Bussière, F. (2014). Direct splash dispersal prevails over indirect and subsequent spread during rains in *Colletotrichum gloeosporioides* infecting yams. *PLoS One* 9(12):e115757. Perondi, D., Fraisse, C.W., Dewdney, M.M., Cerbaro, V.A., Andreis, J.H.D., Gama, A.B., Junior, G.J.S., Amorim, L., Pavan, W. and Peres, N.A. (2020). Citrus advisory system: A web-based postbloom fruit drop disease alert system. *Computers and electronics in agriculture* 178:105781.

Porras, M.F., Navas, C.A., Agudelo-Cantero, G.A., Santiago-Martínez, M.G., Loeschcke, V., Sørensen, J.G., Crandall, S.G., Biddinger, D. and Rajotte, E.G. (2023). Extreme heat alters the performance of hosts and pathogen. *Frontiers in Ecology and Evolution* 11:1186452.

Pritchard SG, Rogers HH, Prior SA, Peterson CM (1999) Elevated CO₂ and plant structure: a review. *Global Change Biology* 5:807–837.

Oiu, J., Liu, Z., Xie, J., Lan, B., Shen, Z., Shi, H., Lin, F., Shen, X. and Kou, Y. (2022). Dual impact of ambient humidity on the virulence of Magnaporthe and basal orvzae resistance in rice. *Plant*, Cell Å Environment 45(12):3399-3411.

Sapoukhina, N., Boureau, T. and Rousseau, D. (2022). Plant disease symptom segmentation in chlorophyll fluorescence imaging with a synthetic dataset. *Frontiers* in *Plant Science* 13:969205.

Sathaye, J., Shukla, P.R. and Ravindranath, N.H. (2006). Climate change, sustainable development and India: Global and national concerns. *Current science* 90(3):314-325.

Savary, S., Willocquet, L., Pethybridge, S.J., Esker, P., McRoberts, N. and Nelson, A. (2019). The global burden of pathogens and pests on major food crops. *Nature* ecology & evolution 3(3):430-439. Saxony, L., Richerzhagen, D., Racca, P., Zeuner, T., Kuhn, C., Falke, K., Kleinhenz, B. and Hau, B. (2011). Impact of climate change on the temporal and regional occurrence of *Cercospora* leaf spot in Lower Saxony. *Journal of Plant Diseases and Protection* 118:168-177.

Schroth, G., Krauss, U., Gasparotto, L.J.A.D., Duarte Aguilar, J.A. and Vohland, K. (2000). Pests and diseases in agroforestry systems of the humid tropics. *Agroforestry* systems 50:199-241.

Singh, B.K., Delgado-Baquerizo, M., Egidi, E., Guirado, E., Leach, J.E., Liu, H. and Trivedi, P. (2023). Climate change impacts on plant pathogens, food security and paths forward. *Nature Reviews Microbiology* 21(10):640-656.

Slade, A.J. and Knauf, V.C. (2005). TILLING moves beyond functional genomics into crop improvement. *Transgenic research* 14:109-115.

Smith, P., Martino, D., Cai, Z., Gwary, D., Janzen, H., Kumar, P., McCarl, B., Ogle, S., O'Mara, F., Rice, C. and Scholes, B. (2008). Greenhouse gas mitigation in agriculture. *Philosophical transactions of the royal Society B: Biological Sciences* 363(1492):789-813.

Stefansson, T.S., McDonald, B.A. and Willi, Y. (2013). Local adaptation and evolutionary potential along a temperature gradient in the fungal pathogen *Rhynchosporium commune. Evolutionary Applications* 6(3):524-534.

Tada, T., Tanaka, C., Katsube-Tanaka, T. and Shiraiwa, T. (2021). Effects of wounding and relative humidity on the incidence of Phytophthora root and stem rot in soybean seedlings. *Physiological and Molecular Plant Pathology* 116:101737.

Tadesse, W., Sanchez-Garcia, M., Assefa, S.G., Amri, A., Bishaw, Z., Ogbonnaya, F.C. and Baum, M. (2019). Genetic gains in wheat breeding and its role in feeding the world. *Crop Breeding*, *Genetics* and *Genomics* 1(1):e190005.

Tiwari, I.M., Jesuraj, A., Kamboj, R., Devanna, B.N., Botella, J.R. and Sharma, T.R. (2017). Host delivered RNAi, an efficient approach to increase rice resistance to sheath blight pathogen (*Rhizoctonia solani*). *Scientific reports* 7(1):7521.

Toniutti, L., Breitler, J.C., Etienne, H., Campa, C., Doulbeau, S., Urban, L., Lambot, C., Pinilla, J.C.H. and Bertrand, B. (2017). Influence of environmental conditions and genetic background of arabica coffee (*C. arabica* L) on leaf rust (*Hemileia vastatrix*) pathogenesis. *Frontiers in Plant Science* 8:2025.

Váry, Z., Mullins, E., McElwain, J.C. and Doohan, F.M. (2015). The severity of wheat diseases increases when plants and pathogens are acclimatized to elevated carbon dioxide. *Global change biology* 21(7):2661-2669.

Velásquez, A.C., Castroverde, C.D.M. and He, S.Y. (2018). Plant–pathogen warfare under changing climate conditions. *Current biology* 28(10):R619-R634.

World Meteorological Organization. (2024). Greenhouse gas concentrations surge again to new record in 2023, Available from https://wmo.int/media/news/greenhous e-gas-concentrations-surge-again-newrecord-2023 [18 November 2024].

Yao, L., Jiang, Z., Wang, Y., Hu, Y., Hao, G., Zhong, W., Wan, S. and Xin, X.F. (2023). High air humidity dampens salicylic acid pathway and NPR1 function to promote plant disease. *The EMBO Journal* 42(21):e113499.

Zayan, S.A. (2019). Impact of climate change on plant diseases and IPM strategies. In *Plant Diseases-Current Threats and Management Trends*. IntechOpen.

Zhai, Z., Martínez, J.F., Beltran, V. and Martínez, N.L. (2020). Decision support systems for agriculture 4.0: Survey and challenges. *Computers and Electronics in Agriculture* 170:105256. Zhou, Y., Van Leeuwen, S.K., Pieterse, C.M., Bakker, P.A. and Van Wees, S.C. (2019). Effect of atmospheric CO₂ on plant defense against leaf and root pathogens of Arabidopsis. *European Journal of Plant Pathology* 154:31-42.